PREVALENCE OF VANCOMYCIN RESISTANT ENTEROCOCCUS AND ITS

ANTTIBIOTIC RESISTANCE PATTERN IN PATIENTS ADMITTED IN

TERTIARY CARE HOSPITAL

Shadma Yaqoob¹, Priyanka Shukla², Fareya Haider³, Zahida Parveen Dar⁴, Vaibhav Shukla⁵

¹ Assistant Professor, ² Lecturer, ^{3, 4} Junior Resident, Department of Microbiology, ⁵ Associate

Professor, Department of Medicine, Era's Lucknow Medical College, Lucknow – UP (India).

Correspondence:drshadmayagoob@yahoo.com

Abstract

Enterococci are indigenous flora of the intestinal tract, oral cavity & genitourinary tract of human & are important opportunistic pathogens, especially in hospitalized patients. This genus is resistant to many antimicrobial agents commonly used in hospitals including β-lactam antibiotics, glycopeptides aminoglycosides. In recent yrs the incidence of enterococcal infection has increased making the second most common nosocomial pathogen. The present study reveals the problem of mutidrug resistant enterococci and emergence of VRE. Emergence of high-level resistance to aminoglycosides (HLAR), βlactam antibiotics and to vancomycin by some strains, with multidrug resistance has led to the failure of synergistic effects of combination therapy, more often in hospitalized patients and previously treated with antibiotics.

Key words: Nosocomial infection, Vancomycin resistant enterococcus, Polymicrobial infection.

Introduction

Enterococci are indigenous flora of the intestinal tract, oral cavity & genitourinary tract of human & are important opportunistic

pathogens, especially in hospitalized patients.1

E.faecalis(80-90%) & E.faecium(5-10%) are

two commonly prevalent species which are

human pathogens capable of causing urinary

tract infections, intra-abdominal infections, pelvic wound infections, biliary tract infections, respiratory infections. neonatal sepsis accompanied by bacteremia or meningitis or both2. Other enterococcal species are identified less often. This genus is resistant to number of antimicrobial agents commonly used in hospitals including B-lactam antibiotics. glycopeptides and aminoglycosides. They can also rapidly express resistance to many antibiotics by acquisition of plasmids & transposable elements3. In recent yrs the incidence of enterococcal infection has increased, making it the second most common nosocomial pathogen reported to the National Nosocomial Infection surveillance system4. However emergence of high-level resistance to aminoglycosides (HLAR), β-lactam antibiotics and to vancomycin by some strains, with multidrug resistance has led to the failure of synergistic effects of combination therapy, more often in hospitalized patients previously treated with antibiotics.5

Material & Method:

This observational study was conducted after approval from institutional ethical committee in Department of Microbiology, the Era's Lucknow Medical College and Hospital, Lucknow indoor patients among from December 2007 to December 2008. Eighty six enterococcus strains were isolated from various clinical specimens including pus, urine, wound swab, catheters, blood, sputum, throat swab, cerebrospinal fluid, high vaginal swab and other body fluids, collected from patients of all age group admitted in the Departments of Surgery, Gynecology and Obstetrics, Medicine, Pediatrics and Orthopedics. Past history of the patients was recorded for diabetes mellitus. chronic renal illness and other chronic illness leading to prolonged hospitalization.

Specimen Processing was Done in Two Parts

Part I- Isolation and identification of Enterococcus by culture and biochemical tests.

Culture of Specimens

All the specimens received in the bacteriology laboratory were inoculated on Blood agar and

McConkey agar plates & incubated at $37\Box C$ for 24-48 hours.

Identification and speciation of Enterococcus

Presumptive identification was done on the basis of colony characteristics, Gram's staining, catalase test. Confirmation was done by growth in 6.5% NaCl, bile aesculin hydrolysis, Production of acetoin, Pyruvate utilization, Arginine decarboxylation, Haemolysin production, Tellurite reduction.

Part II- In Vitro Antibiotic Susceptibility Testing by Disc Diffusion Method of Kirby Bauer and Minimum Inhibitory Concentration(MIC) by Agar Dillution Method. Disc testing was performed according to the CLSI guidelines. Muller-Hinton agar was used as media. It was inoculated with a suspension of each organism equivalent to 0.5 McFarland turbidity standard and discs were applied. Inhibition zones were interpreted according to CLSI guidelines. Control strains used were E faecalis ATCC-29212(susceptible) and E.faecalis ATCC-51299(resistant).

Disk Diffusion Sensitivity by Vancomycin

30 microgram disc was carried out using Kirby Bauer method. Muller-Hinton agar (MHA) plates were overlaid with the inoculums turbidity equivalent to that of a 0.5 McFarland Standard following CLSI criteria. Zone diameters were measured at 24 hrs following CLSI criteria i.e. zone of inhibition more than or equal to 17 mm reported as sensitive and less than 17mm were further confirmed by growth on BHI(Brain Heart Infusion) screen agar and MIC testing.

Determination of Vancomycin MIC was done by Agar Dilution Method. Enterococcus strains that were resistant and intermediate sensitive by Disk Diffusion method were further tested by BHI Vancomycin Screen Agar. As per CDC guidelines, in-house prepared BHI agar (Hi-Media, India) screen plates containing 6 microgram/ml Vancomycin (Lilly Pharma, Giessen, Germany) was prepared. Inoculum suspensions were prepared by selecting colonies from overnight growth on nutrient agar plates. The colonies were transferred to sterile saline to produce a suspension that

matched the turbidity of 0.5 McFarland Table andard. The final inoculum concentration of 105 to 106 CFU per spot was prepared by adding the sterile saline to the bacterial suspension. These suspensions were spot inoculated on BHI screen agar plates and plates were incubated for 24 hrs. at 35°C aerobically. Any visible growth was indicated for Vancomycin resistance.

Further detection of VRE was done by MIC by vancomycin agar dilution method using MHA. The concentrations tested ranged from 2 μ g/ml to 1024 μ g/ml of vancomycin.

Results:

Eighty six Enterococcus strains were isolated after identification by standard biochemical tests. All were tested for Vancomycin resistance using Mueller Hinton agar by Kirby-Bauer disc diffusion method 6,7. six isolates showed resistance and six were intermediate sensitive after 18-24 hours incubation at 35°C.

Table 1- Distribution of *enterococcus* strains isolated from the various clinical specimens

Samples	No.of samples	No. of Enterococci		
Urine	1900	58		
Blood	290	5		
Pus	380	10		
Vaginal swab	180	8		
Throat swab, Pleural fluid, Ascitic fluid, Peritonial fluid Bile, CSF, tissue aspirate	365	5		
Total	3115	86		

Table 2- MIC of the VRE isolated (n=6)

Wards	Species	Source of	Sensitivity	MIC by	
		Specimen	pattern by	broth	
			disc method	dilutionmethod	
ICU	E. faecalis	Urine	Resistant	512 μg/ml	
ICU	E. faecalis	Urine	Resistant	64μ g/ml	
GYNAE	E. faeicum	Urine	Resistant	64μ g/ml	
GYNAE	E. faecalis	Pus	Resistant	32μ g/ml	
POST- OP	E. faecalis	Urine	Resistant	16μ g/ml	
ICU	E. faecalis	Urine	Resistant	16 μg/ml	

Enterococcus strains that were resistant and intermediate sensitive by Disk Diffusion method were further tested by BHI Vancomycin Screen Agar. Vancomycin screen agar showed only 6 resistant strains and the other 6 intermediate strains became susceptible after screening. MIC also showed concordant results with Vancomycin screen agar ie. the percentage of sensitive strains had increased from 86.04% to 93.02%. The MIC of VRE ranged from 16 - 512μgm/ml. All 6 VRE isolates were further tested for different antibiotics susceptibility and resistance pattern.8

Table 3- Resistance pattern of VRE strains among the patients

VAN	Amp	HSG	Nx	Ср	Е	Te	Lz	Pm
MIC								
μgm/ml								
16 μg	R	R	S	R	R	S	S	R
16 μg	R	S	R	R	R	S	S	S
32 μg	R	R	R	R	R	R	S	R
64 μg	R	R	R	R	R	S	S	S
64 μg	R	S	R	R	R	S	S	S
512 μg	R	R	R	R	R	S	S	R

(Van-Vancomycin, Amp-Ampicillin, HSG-High strength Gentamycin, Nx- Norfloxacin, Cp-Ciprofloxacin, E-Erythromycin, Te-Teicoplanin, Lz-Linezolid, Pm-Pristinomycin)

One of the VRE strain was found to be resistant to teicoplanin and remaining 5 were sensitive. This drug is rapidly and completely absorbed after oral administration with a mean bioavailability of approximately 100%9. We observed that this drug

is effective against multiple drug resistant strains of Enterococcus.

Discussion

Out of 86 isolates, 75 were isolated in pure culture while the rest 11 were in combination with other bacteria ie: 12.7% of enterococcal infections were polymicrobial10. It suggests that enterococci can act synergistically with other intestinal bacteria to enhance the rate of infection.

The MIC of VRE ranged from 16 - 512µgm/ml. Chaudhary U et al. 200711, had also reported approximate result. Out of 6 VRE, five resistant isolates were from urine specimen and one from pus that was approximately similar to Taneja et al. (2004) from PGI Chandigarh. All were E.faecalis species except one that was E.faecium. All 6 VRE were isolated from admitted patients. Of 6 VRE isolates, 4 were female patients, 2 in post operative ward, 2 in Gynae ward and 2 were males - 1 in I.C.U. and 1 in male medicine ward and were associated with urinary tract infection and catheterization with prolonged hospital stay.

One of the VRE strain was found to be resistant to teicoplanin and remaining 5 were sensitive so this study had given the conclusion that isolate that was

VRE and teicoplanin resistant was phenotype VanA and the remaining 5 isolates which were teicoplanin sensitive, probably VanB or any other phenotypes12. An indoor female patient who was suffering from UTI with septicaemia and the isolate was E.faecalis which was resistant to all drugs except Linezolid but she did not respond to the treatment& died during hospital stay only.

Conclusion: An important feature in the emergence of the enterococci as a cause of nosocomial infection is their increasing resistance to a wide range of antibiotics and their ability to acquire resistance to all currently available antibiotics either by mutation or by receipt of foreign genetic material through the transfer of plasmids and transposons13. High Level Aminoglycoside Resistant strains with multidrug resistance has led to the failure of synergistic effects of combination therapy14,15. The present study reveals the problem of mutidrug resistant enterococci and emergence of VRE. Thus we suggest to promote more the rational use of antibiotics in health care settings, more surveillance studies in order to monitor changes in enterococcal resistance patterns.

So a good antibiotic policy should be laid down between the clinician and microbiologist in all tertiary care hospitals and a strict antibiotic regimen should be applied by clinicians. Those patients identified with history of chronic illness like diabetes mellitus, renal failure, peritoneal dialysis should be dealt with utmost care.

Competing Interest: None declared

Funding: Nil

References

- Hancock LE and Gilanome MS, 2000,
 Pathogenicity of enterococci, p. 251-258
 in V.A. Fischetti, R. R. Novick, J.J.
 Ferretti, D.A. Portnoy and J.I. Rood (ed),
 Gram-Positive pathogens ASM Press,
 Washington, D.C.
- Felmingham D, Wilson. Enterococcus species in urinary tract infection. Clin infect Dis.1992; 15: 295-301.
- 3. Mohanty S, Jose S, Singhal R, sood S, Dhawan B, Dask BK. Species prevalence and antimicrobial susceptibility of enterococci isolated in a tertiary care hospital of north India. Southeast Asian J Trop Med public Health 2005; 36: 962-5.

- Centre of Disease Control, Nosocomial enterococci resistant to vancomycin United States. National Nosocomial infection surveillance MMWR 1993; 42
- Ghoshal U, Garg A, Tiwari DP, Ayyagari A.Emerging vancomycin resistance in enterococci in India. Indian J Pathol Microbial 2006; 49:620-2.
- Desai PJ, pandit D, Mathur M, Gogute A.
 Prevalence, identification and distribution of various species of enterococci isolated from clinical specimen. Ind J Med Microb 2001; 19: 132-137.
- Edet E Udo, Noura AI-Sweih, Oludotan
 A, Phillips and Tulsi D Chug. Species
 prevalence and antibacterial resistance of
 enterococci. Med Microbiol 2003; 52:
 163-168.
- 8. Sood S, Malhotra M, Das B K, Kapil A. Enterococcal infections and antimicrobial resistance.Indian J Med Res 2008; 128: 111-121.

- 9. EI-Khoury J, Fishman JA. Transpl Infect
 Dis. 2003 sep; 5 (3): 121-5
- 10. Chaudhary U, et al J Infect Dis
 Antimicrob Agents 2007;24:55-62
- 11. Chaudhary U, Shamma M, Yadav.

 Antimicrobial susceptibility pattern of common and unusual Enterococcus from clinical specimens.J Infect Dis Microbial Agents 2007; 24: 55-62.
- 12. H.S.Gold. Antimicrobial resistance. CID2001:33(15 July).219
- 13. Bhat KG, Paul C, Anantha Krishna NC.Drug resistant enterococci in aSouth Indian hospital. Trop Doct1998; 28: 106-7.
- 14. Agarwal JB, Kalyan R, Singh M. High-Level Aminoglycosides Resistance and β-Lactamase Production in Enterococci at a Tertiary Care Hospital in India. Jpn J infect Dis 2009; 62: 158-159.597-599.
- 15. Mendiratta D K, Kaur H,

 Deotale V, Thamke D C, Narang R,

Narang P. Status of High Level
Aminoglycosides Resistance
Enterococcus faecium and Enterococcus
faecalis in rular Hospital of central India.
Indian J Med Microbial 2008; 26: 369371

Figures showing results of MIC of Vancomycin



